

Predicting the side-chain dihedral angle distributions of nonpolar, aromatic, and polar amino acids using hard sphere models

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ABSTRACT

The side-chain dihedral angle distributions of all amino acids have been measured from myriad high-resolution protein crystal structures. However, we do not yet know the dominant interactions that determine these distributions. Here, we explore to what extent the defining features of the side-chain dihedral angle distributions of different amino acids can be captured by a simple physical model. We find that a hard-sphere model for a dipeptide mimetic that includes only steric interactions plus stereochemical constraints is able to recapitulate the key features of the back-bone dependent observed amino acid side-chain dihedral angle distributions of Ser, Cys, Thr, Val, Ile, Leu, Phe, Tyr, and Trp. We find that for certain amino acids, performing the calculations with the amino acid of interest in the central position of a short α -helical segment improves the match between the predicted and observed distributions. We also identify the atomic interactions that give rise to the differences between the predicted distributions for the hard-sphere model of the dipeptide and that of the α helical segment. Finally, we point out a case where the hard-sphere plus stereochemical constraint model is insufficient to recapitulate the observed side-chain dihedral angle distribution, namely the distribution $P(\chi_3)$ for Met.

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INTRODUCTION

Many types of interactions, including purely repulsive steric, electrostatic, and the attractive component of van der Waals interactions, determine the structure of proteins in general and the conformations of amino acid side chains in particular. However, the relative contributions of the different types of interactions are not known. For example, the dominant interactions that control the side-chain dihedral angle distributions for each residue have not been identified, even though there is now a wealth of highresolution structural data on thousands of proteins. We seek to determine to what extent the key features of the side-chain dihedral angle distributions of different amino acids observed in protein crystal structures can be captured by steric repulsion and stereochemical constraints alone.

A similar question motivated Ramachandran and colleagues to model peptides as hard spheres with stereochemical constraints for the bond lengths and angles.^{1,2} Using such hard-sphere calculations, they identified the backbone dihedral angle combinations (φ and ψ) for an alanyl dipeptide mimetic that are sterically allowed, given physically reasonable values for the atom sizes, bond lengths, and bond angles. They found that large regions of φ and ψ space are disallowed due to steric clashes between atoms in the dipeptide. Moreover, we now know that essentially every amino acid in every protein

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structure obeys the hard-sphere limits on ϕ and ψ defined by Ramachandran and coworkers. 3,4

Dunbrack and colleagues have also gained significant insight into the side-chain conformations by analyzing high-resolution protein crystal structures.^{5–8} They emphasized that the side-chain dihedral angle distributions are rotameric, with high probabilities at specific χ_1 and χ_2 combinations that depend sensitively on the backbone dihedral angles. They also showed that certain rotamers are rare because of steric repulsions analogous to those that constrain the conformations of hydrocarbon chains. However, these studies did not specifically address whether hard-sphere interactions plus stereo-chemical constraints alone can recapitulate the key features of side-chain dihedral angle distributions.

Common computational strategies for calculating the side-chain dihedral angle distributions in proteins involve quantum mechanical calculations⁹ and molecular mechanics force fields, such as Amber,¹⁰ CHARMM,¹¹ GROMOS,¹² and OPLS.¹³ However, these simulation methods require prohibitively large computing resources when considering large proteins. In addition, molecular mechanics force fields directly sample the experimentally measured backbone and side-chain dihedral angle distributions using knowledge-based potentials, such as CHARMM-CMAP¹⁴ and χ -CMAP,¹⁵ or Amber-ILDN.¹⁶ Thus, the molecular mechanics force fields do not allow one to quantify the distinct contribution of steric repulsive interactions to the side-chain dihedral angle distributions and build-in the observed side-chain dihedral angle distributions that we seek to predict.

In this manuscript, we pose the key question: To what extent, do steric interactions plus stereochemical constraints determine the side-chain dihedral angle distributions for each residue? Specifically, we determine the sterically allowed side-chain dihedral angle distributions for the nonpolar (Leu, Ile, and Val), aromatic (Phe, Tyr, and Trp), and polar residues (Cys, Ser, and Thr) in either α -helix or β -sheet backbone conformations using a hardsphere model that only includes steric interactions plus stereochemical constraints for the bond lengths and angles. For each residue, we determine which features of the side-chain dihedral angle distributions can be explained by a hard-sphere model, in the context of an amino acid dipeptide mimetic (Fig. 1). For several amino acids, the major features of the observed distributions are recapitulated by the hard-sphere model for a dipeptide mimetic. For certain other amino acids, some features of the observed distributions are not captured by the hard-sphere model for a dipeptide mimetic. These discrepancies were especially apparent for α -helical backbone conformations. We therefore also performed hardsphere calculations of side-chain dihedral angle distributions for each amino acid in the context of a nineresidue α -helical segment. In some cases, it is clear that certain side-chain dihedral angle combinations are sterically disallowed due to atomic clashes with the surrounding residues in the α -helical segment, not by clashes within the local environment of the dipeptide mimetic. The ability to predict amino acid side-chain dihedral angle distributions using a simple physical model provides an efficient tool for computational protein design strategies that does not rely on knowledge-based potentials for determining side-chain dihedral angles.¹⁷

MATERIALS AND METHODS

Databases of protein crystal structures

Our calculations of the sterically allowed side-chain dihedral angle distributions are compared with the observed side-chain dihedral angle distributions obtained from a database of high-resolution protein crystal structures provided by Dr. Roland Dunbrack, Jr.¹⁸ This database is composed of 850 non-homologous protein structures with resolution 1.7 Å or less, side chain B-factors per residue 30 Å² or less, and R-factors 0.2 or less.

We used a higher-resolution (1.0 Å or less) set of 221 structures⁸ to fix the bond lengths, bond angles, and ω backbone dihedral angles of the dipeptide mimetics for our calculations. The higher-resolution set limits the refinement bias on the bond lengths, bond angles, and ω dihedral angles associated with the crystal structure determination, but there are too few structures in this set to provide meaningful comparisons with the calculated side-chain dihedral angle distributions. The numbers of each residue type in both databases are displayed in Table S1 in the Supporting Information. In addition, we identified 53 protein crystal structures containing Nle in the protein data bank (PDB). 24 of the 53 crystal structures are nonhomologous. After removing structures with resolution higher than 1.7 Å, 11 Nle amino acids remained.

Hard-sphere plus stereochemical constraint model of dipeptide mimetics and α-helical segments

Figure 1 shows a stick representation of the Ala dipeptide mimetic as well as nine other residues (Val, Thr, Phe, Ser, Cys, Tyr, Leu, Ile, and Trp). The φ backbone dihedral angle is defined by the clockwise rotation around the N-C_{\alpha} bond (viewed from N to C_{\alpha}) involving the backbone atoms C-N-C_{\alpha}-C. The ψ backbone dihedral angle is defined by the clockwise rotation about the C_{\alpha}-C bond (viewed from C_{\alpha} to C) involving the backbone atoms N-C_{\alpha}-C-N. The definitions of the ω backbone dihedral angles and the side-chain dihedral angles χ_i of the nine residues considered in this work are provided in Table S2 in Supporting Information. All dihedral angles listed in the Table S2 in Supporting



Stick representation of dipeptide mimetics. Stick representation of dipeptide mimetics for nine residues: Val, Thr, Ser, Cys, Leu, Ile, Phe, Tyr, and Trp. The side-chain dihedral angles χ_1 and χ_2 are highlighted, with positive angles indicated by the arrows. Carbon, nitrogen, oxygen, hydrogen, and sulfur atoms are shaded pink, blue, red, white, and yellow, respectively. We also include the Ala dipeptide mimetic to label the backbone atoms and define the backbone dihedral angles (φ and ψ). Atoms with superscripts i - 1 and i + 1 refer to the residue order relative to the central residue *i*.

Information range from 0° to 360°, except χ_2 for the aromatic residues Tyr, Trp, and Phe, which range from 0° to 180°. We show in the Supporting Information that the side-chain dihedral angle distributions for Tyr, Trp, and Phe are similar for 0° < χ_2 < 180° and 180° < χ_2 < 360° because of the approximate ring inversion symmetry.

We constructed hard-sphere representations of dipeptide mimetics and α -helical segments for each of the nine residues. The structure of the dipeptide mimetics is N-acetyl-X-N'-methylamide, where X is one of the side chains of the nine residues we studied (Fig. 1). Particular combinations of the bond lengths, bond angles, and ω backbone dihedral angles for each dipeptide mimetic were obtained from the ≤ 1.0 Å subset of protein crystal structures. The atomic radii were set to 1.05 Å for Hydrogen, 1.5 Å for sp³ Carbon, 1.4 Å for sp² Carbon, 1.4 Å for Nitrogen, and 1.45 Å for Oxygen, based on our previous studies.^{19–21} We fixed the radius of Sulfur to be 1.8 Å consistent with many prior studies^{22–30} and

the mean bond lengths $\mu = 1.804 \pm 0.015$ and 1.792 ± 0.015 for C_{γ} -S_{δ} and C_{ε} -S_{δ} in Met from the 1.0 Å database. Hydrogen atoms were added to the structures using the REDUCE software package,³¹ which places hydrogen atoms based on their nuclear positions.

As shown in Figure 5, the α -helical segments contain nine residues with the target residue positioned in the center at location *i*, and the side-chains of the other eight residues in the segment have been removed. The α helical segments extracted from the 1.0 Å set of structures satisfy the following criteria: (1) no missing atoms in the backbone of the segment, (2) no Pro residues, and (3) all backbone dihedral angle combinations possess $-80^{\circ} < \phi < -20^{\circ}$ and $-65^{\circ} < \psi < -20^{\circ}$. In contrast to our previous studies,^{19,20,32} we do not change the backbone dihedral angles of the dipeptide mimetics and α helical segments that were extracted from the highresolution database.

Calculation of the probability distribution of sterically allowed side-chain dihedral angles

Given a particular dipeptide mimetic or α -helical segment, we rotate the side chain of the target residue to a particular conformation specified by $\{\chi_1, \chi_2, \ldots\}$ in small increments $\Delta \chi = 5^\circ$. For each side-chain conformation, we determine the separation r_{ij} between the centers of all pairs of nonbonded atoms *i* and *j* (with both atoms located on the side chain or with one on the side chain and the other on the backbone). If the separation r_{ij} between all nonbonded atom pairs for a conformation satisfies $r_{ij} \geq \sigma_{ij}$, where σ_{ij} is the sum of the radii of atoms *i* and *j*, this conformation is sterically allowed. We then perform this calculation for all possible side chain conformations for each dipeptide mimetic or α -helical segment.

To calculate the probability distributions of sterically allowed side-chain dihedral angles, $P(\chi_1)$ for Val, Thr, Ser, and Cys, and $P(\chi_1, \chi_2)$ for Leu, Ile, Phe, Tyr, and Trp in a dipeptide mimetic or α -helical segment, we first count the number of sterically allowed side-chain dihedral angle combinations in each 5° bin or 5° × 5° box, and then sum over all residues selected from the highresolution Dunbrack database. Summing over structures in the Dunbrack database allows us to average over random bond length, bond angle, and ω dihedral angle fluctuations.

To investigate the dependence of the side-chain dihedral angle distributions on the backbone conformation, we identified dipeptide mimetics with φ and ψ within $\pm 10^{\circ}$ of canonical α -helix ($\varphi = -57^{\circ}$, $\psi = -47^{\circ}$) or β sheet ($\varphi = -119^{\circ}$, $\psi = 113^{\circ}$) values, and calculated the probability distributions for the side-chain dihedral angles for α -helical and β -sheet backbone conformations separately. We normalize the distributions so that $\int P(\chi_1) d\chi_1 = 1$ or $\int P(\chi_1, \chi_2) d\chi_1 d\chi_2 = 1$.

RESULTS

We seek to develop a predictive understanding of the side-chain dihedral angle distributions of several classes of amino acids. We employ a simple approach: modeling amino acids as hard-spheres with stereochemical constraints on the bond lengths and angles. Briefly, using Val for illustration, our strategy is as follows. We first identify all occurrences of the amino acid of interest in the 1.0 Å Dunbrack database of protein crystal structures. (See Materials and Methods for a description of the Dunbrack database of high-resolution protein crystal structures.) For example, there are 424 Val with α -helical φ - ψ combinations in the database as shown in Table S1 in Supporting Information. Each of these Val residues possesses slightly different bond lengths and angles. We fix the original backbone dihedral angles φ and ψ , and for each of these Val residues, we calculate whether or not a particular side-chain dihedral angle, χ_1 , is sterically allowed or disallowed. We then sum these results for all 424 Val with α -helical backbone conformations to obtain a probability distribution $P(\chi_1)$ for each χ_1 for Val (α helix). We compare the predicted side-chain dihedral angle distributions to the observed distributions obtained from the 1.7 Å (not 1.0 Å) Dunbrack database to have sufficient data (850 vs. 221 protein structures) to calculate the probability distribution for χ_1 . We follow a similar procedure for Ser, Cys, Thr, Ile, Leu, Phe, Tyr, and Trp.

To improve the agreement between our predictions of the side-chain dihedral angle distributions and the observed distributions for certain amino acids, we performed hard-sphere calculations with the target residue positioned in the center of a nine-residue α -helical segment and the side-chains of the other eight residues in the segment removed (i.e., only the backbone remains). See Materials and Methods for a more complete description of the computational methods we employ.

Below, we organize the results into "amino acid types" for clarity. Specifically, we group Ser and Cys; Val and Thr; Ile and Leu; and then Phe, Tyr, and Trp. For each residue, we calculate the side-chain dihedral angle distributions for both α -helical and β -sheet backbone conformations for a dipeptide mimetic as described for Val above. We discuss which features of the observed sidechain dihedral distributions our hard-model can reproduce, and which features it cannot. In several cases, we also compare the results for the side-chain dihedral angle distribution obtained from the dipeptide mimetic with calculations for the same amino acid in the center of a nine-residue α -helical segment.



Side-chain dihedral angle distributions for Ser and Cys dipeptide mimetics: Comparison of the observed (dotted red lines) and calculated (solid blue lines) probability distributions $P(\chi_1)$ of the side-chain dihedral angle χ_1 for Ser and Cys in dipeptide mimetics with backbone dihedral angles φ and ψ within $\pm 10^\circ$ of canonical α -helix ($\varphi = -57^\circ$, $\psi = -47^\circ$) and β -sheet ($\varphi = -119^\circ$, $\psi = 113^\circ$) values. The probabilities are normalized such that $\int P(\chi_1) d\chi_1 = 1$. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

Cys and Ser

In Figure 2, we compare the results of our hard-sphere calculations of the side-chain dihedral angle distributions $P(\chi_1)$ for Ser and Cys in a dipeptide mimetic to the observed distributions for both α -helical and β -sheet backbone conformations. By considering rotations about a single carbon bond in ethane, one would expect three highly probable conformations at $\chi_1 = 60^\circ$, 180° , and 300° . The C_{α} - C_{β} bonds in Cys and Ser are analogous to the carbon-carbon bond in ethane, but Cys and Ser have asymmetric substituents attached to C_{α} and C_{β} . The observed distribution for Ser (α -helix) shows three peaks at $\chi_1 = 60^\circ$, 180° , and 300° . However, the peak at $\chi_1 = 60^\circ$ is greatly diminished for Cys (α -helix) and nearly absent for both Ser and Cys (β -sheet). Our pre-

dictions from the hard-sphere model for a dipeptide mimetic show strong similarities to the observed sidechain distributions for both Ser and Cys in both α -helix and β -sheet backbone conformations. (See Fig. S2 in Supporting Information for estimates of the error bars for the predicted and observed side-chain dihedral angle distributions.) The hard-sphere model predicts that $\chi_1 = 60^\circ$ is more strongly disfavored for Ser (β -sheet) compared to Ser (α -helix) and $\chi_1 = 60^\circ$ is strongly disfavored for Cys (both α -helix and β -sheet).

We identified the steric clashes that determine the form of the allowed side-chain dihedral angle distributions for Cys and Ser (Fig. S1 in Supporting Information). We tracked how the separations between the 47 pairs of atoms change as we vary χ_1 for both Cys and Ser dipeptides in both α -helix and β -sheet backbone conformations. For each amino acid in each backbone conformation, we studied dipeptide models that sample bond length and bond angle combinations from the 1.0 Å database. Clashes between the side-chain O_{γ} and backbone N^{i+1} and H^{i+1} atoms (where H^{i+1} is the hydrogen on N^{i+1}) disallow 50% of the Ser dipeptides at $\chi_1 = 60^\circ$. Similar calculations show that clashes between the O_{γ} -O, S_{γ} -N^{*i*+1}, and S_{γ} -H^{*i*+1} atom pairs disallow nearly all Ser ($\dot{\beta}$ -sheet) and Cys (α -helix) residues at $\chi_1 = 60^\circ$. In contrast, nearly all Cys (α -helix) residues at $\chi_1 = 180^\circ$ are sterically allowed.

Val and Thr

As with Cys and Ser, one might expect three highly populated side-chain conformations near $\chi_1 = 60^\circ$, 180° , and 300° for Val and Thr. Indeed, three peaks are observed in the probability distributions $P(\chi_1)$ for Val and Thr (α -helix), but not for Val and Thr (β -sheet). The calculated $P(\chi_1)$ for the β -sheet conformations of both the Val and Thr dipeptide mimetics are similar to the observed $P(\chi_1)$ [Fig. 3(c,f)], *i.e.* we find one strong peak at $\chi_1 = 180^\circ$ for Val and at $\chi_1 = 300^\circ$ for Thr.¹⁹ Note that in the IUPAC nomenclature³³ the definition of χ_1 is different for Thr and Val with respect to the methyl branch. Therefore, $\chi_1 = 180^\circ$ in Val is equivalent to $\chi_1 = 300^\circ$ in Thr.

Because the hard-sphere model for the dipeptide mimetic overpredicts the peaks at $\chi_1 = 60^\circ$ and 300° for Val (α -helix) as well as the peaks at $\chi_1 = 60^\circ$ and 180° for Thr (α -helix) as shown in Figure 3(a,d), we repeated the calculation in a short α -helical segment to determine if inter-residue interactions were influencing the observed distributions. The calculated P(χ_1) from the hard-sphere model for Val and Thr in α -helical segments better corresponds with the observed distributions. In particular, the calculated peaks at $\chi_1 = 300^\circ$ for Val and $\chi_1 = 60^\circ$ and 180° for Thr are reduced, as shown in Figure 3. The probability near $\chi_1 = 300^\circ$ for Val is reduced due to clashes between the side-chain methyl $-C_{\gamma 2}H_3$ group (at



Side-chain dihedral angle distributions for Val and Thr in dipeptide mimetics and α -helical segments: Comparison of the observed (dotted red lines) and calculated (solid blue lines) probability distributions $P(\chi_1)$ for the side chain dihedral angles χ_1 for Val and Thr in dipeptide mimetics with backbone dihedral angles φ and ψ within $\pm 10^\circ$ of either the canonical α -helix ($\varphi = -57^\circ$, $\psi = -47^\circ$) or β -sheet ($\varphi = -119^\circ$, $\psi = 113^\circ$) values (a, c, d, and f) and in α -helical segments (b and e). The probabilities are normalized such that $\int P(\chi_1) d\chi_1 = 1$. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

location i) with the backbone carbonyl group at location i-4. The probability near $\chi_1 = 180^\circ$ for Thr is reduced due to clashes between the side-chain $-C_{\gamma 2}H_3$ group (at location i) with the backbone carbonyl groups at locations i-3 and i-4. The probability near $\chi_1 = 60^\circ$ is reduced by similar clashes with the backbone carbonyl group at location i-4 only.

Leu and lle

Our previous hard-sphere calculations²⁰ of the sidechain dihedral angle distributions $P(\chi_1, \chi_2)$ for Leu and Ile in a dipeptide mimetic capture the main features of the observed distributions for both α -helix and β -sheet backbone conformations (Fig. 4). However, there are a few subtle differences. For example, in the observed distribution for Leu (α -helix) conformations near $\chi_1 = 300^\circ$, $\chi_2 = 300^\circ$ (box 9) are rarely populated, representing only 2% of observed conformations. However, our hard-sphere calculations for Leu (α -helix) in the dipeptide mimetic give 21% for the probability in box 9. In addition, for Ile (α -helix), we predict 25% in box 5 (Fig. 4), whereas the observed probability is 2%.

We therefore also calculated the $P(\chi_1, \chi_2)$ for the hardsphere model of Leu and Ile in an α -helical segment. For Leu, the predicted probability for conformations in box 9 decreases from 21% to 5% [left panels of Fig. 4(b,c)], which more closely matches the observed probability. The decrease in probability in box 9 is caused by clashes between the methyl group $(-C_{\delta 1}H_3)$ of Leu at location i and the backbone carbonyl group (C=O) of the residue at location i-4 (Fig. 5). For Ile, the predicted probabilities for conformations in boxes 5 and 6 change from 18% to 4% and 53% to 78%, respectively [right panels of Fig. 4(b,c)], both of which more closely match the observed probabilities (which are 1% for Box 5 and 85% for Box 6). As shown in Figure 5, the methyl – $C_{\gamma 2}H3$ group of the β branched Ile clashes with the backbone C=O group of both the i-3 and i-4 residues when $\chi_1 < 200^\circ$, which eliminates conformations in box 5.

The observed and calculated probability distributions for the side chain dihedral angles $P(\chi_1, \chi_2)$ for Leu and Ile (β -sheet) are shown in Figure 4. The observed and calculated probability distributions for Leu (β -sheet) both have high probabilities in boxes 6 and 8. However, the hard-sphere model for the Leu dipeptide mimetic (β -sheet) slightly overpredicts conformations in box 9 (11% versus 2%). This slight overprediction is likely because the side chain of Leu at position i would clash with the side chain of the residue at i-2 when it is in a β -sheet structure rather than in a dipeptide mimetic.

For Ile (β -sheet), both the observed and predicted distributions P(χ_1 , χ_2) have a strong peak in box 6 and a minor peak in box 3. The predicted and observed distributions are similar without considering neighboring residues in segments of β -sheet secondary structure, because



 $P(\chi_1, \chi_2)$ for Leu and Ile in dipeptide mimetics and α -helical segments: Comparison of the observed and calculated probability distributions of side-chain dihedral angles $P(\chi_1, \chi_2)$ for Leu (Left column) and Ile (Right column). Row (a): Observed $P(\chi_1, \chi_2)$ for Leu and Ile in α -helical backbone conformations. Row (b): Calculated $P(\chi_1, \chi_2)$ for Leu and Ile dipeptide mimetics in α -helical backbone conformations. Row (c): Calculated $P(\chi_1, \chi_2)$ for Leu and Ile in α -helical segments. Row (d): Observed $P(\chi_1, \chi_2)$ for Leu and Ile in α -helical segments. Row (d): Observed $P(\chi_1, \chi_2)$ for Leu and Ile dipeptide mimetics in β -sheet backbone conformations. Row (e): Calculated $P(\chi_1, \chi_2)$ for Leu and Ile dipeptide mimetics in β -sheet backbone conformations. The probability distributions are normalized such that $\int P(\chi_1, \chi_2) d\chi_1 d\chi_2 = 1$. The probability values, expressed as percentages, within each of the nine (χ_1, χ_2) regions defined by the dotted lines are labeled. The probabilities increase from white to yellow to orange to black. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]



Stick representation of Ile in an α -helical segment: Top row: Stick representation of an α -helical segment with Leu at position i and its sidechain dihedral angles set to (Left) $\chi_1 = 180^\circ$ and $\chi_2 = 60^\circ$ and (Right) $\chi_1 = 300^\circ$ and $\chi_2 = 60^\circ$. The C^{*i*-4} atoms are labeled and atomic clashes between the carbonyl group of residue *i* – 4 and the side-chain of Leu are indicated by semi-transparent spheres. Bottom row: Stick representation of an α -helical segment with Ile at position *i* and its side-chain dihedral angles set to (Left) $\chi_1 = 180^\circ$ and $\chi_2 = 180^\circ$ and (Right) $\chi_1 = 300^\circ$ and $\chi_2 = 180^\circ$. The C^{*i*-4} and C^{*i*-3} atoms are labeled and the atomic clashes between the carbonyl group of residue *i* – 4 and the side chain of Ile are highlighted by semitransparent spheres. Carbon, nitrogen, oxygen, and hydrogen atoms are colored pink, blue, red, and white, respectively. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

Ile is β -branched and clashes between backbone and side-chain atoms within a dipeptide eliminate the (χ_1 , χ_2) combinations equivalent to those in box 9 for Leu.

Phe, Tyr, and Trp

In Figure 6, we compare the calculated dihedral angle distributions $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp dipeptide mimetics in α -helical and β -sheet backbone conformations with the observed distributions. For Phe, Tyr, and Trp in both α -helical and β -sheet backbone conformations the predicted and observed distributions are qualitatively similar. To quantitatively compare the distributions, we decompose (χ_1, χ_2) space into three regions: $0^\circ < \chi_1 < 120^\circ$, $120^\circ < \chi_1 < 240^\circ$, and $240^\circ < \chi_1 < 360^\circ$

with $0^{\circ} < \chi_2 < 180^{\circ}$. For the three aromatic residues, the predicted and observed side-chain dihedral angle distributions agree quantitatively for β -sheet backbone conformations. In contrast, for the three aromatic residues, the hard-sphere dipeptide model overpredicts the probability in the region $240^{\circ} < \chi_1 < 360^{\circ}$ in α -helical backbone conformations by $\approx 20\%$.

In Figure 6, we compare the predicted and observed distributions $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp in the context of the nine-residue α -helical segment. We find that the distributions calculated in this setting more closely match the observed distributions for aromatic residues. In particular, the predicted probabilities in the region $240^\circ\!<\!\chi_1\!<\,360^\circ$ decrease from $\approx~40$ to 20% for Phe and Tyr and from \approx 45 to 30% for Trp, which are all within $\pm 5^{\circ}$ of the values in the observed distributions. We show in Figure 7 that clashes occur between atoms on the aromatic side chain and the carbonyl group of the residue at i-4 when the aromatic residues have $\chi_1 = 300^\circ$ and occur in an α -helical segment. In contrast, these atomic clashes do not occur in the context of a dipeptide mimetic. Based on their examination of the side-chain conformations of Phe in the 61 protein crystal structures available at that time, Sternberg and coworkers³⁴ suggested that such clashes limit the $\chi_1 = 300^\circ$ conformation of Phe, which the current work quantitatively supports.

DISCUSSION AND CONCLUSIONS

We have investigated to what extent the hard-sphere plus stereochemical constraint model can recapitulate the side-chain dihedral angle distributions observed in highresolution protein crystal structures, for Ser, Cys, Val, Thr, Leu, Ile, Phe, Tyr, and Trp. We find that for all of these amino acids, a hard-sphere model of a dipeptide mimetic can explain the main features of the observed side-chain dihedral angle distributions for both α -helix and β -sheet backbone conformations. We also show that in some cases, the match between the predictions from the hard-sphere model and the observed distributions can be improved if we consider the amino acid side chains in the context of an α -helical segment rather than in a dipeptide mimetic.

When considering which amino acids to model using only hard-sphere interactions plus stereochemical constraints, we first chose the non-polar amino acids, because it seemed most likely that their behavior would be well-captured by the hard-sphere model. This proved to be the case for the majority of non-polar residues. The exception is Met (Fig. 8 and Table S1 in Supporting Information). In Figure 9, we compare the observed side-chain dihedral angle distributions, $P(\chi_1)$, $P(\chi_2)$, and $P(\chi_3)$, with those calculated using the hard-sphere model



 $P(\chi_1, \chi_2)$ for a dipeptide mimetic versus α -helical segment for aromatic residues: Comparison of the observed and calculated probability distributions of side-chain dihedral angles $P(\chi_1, \chi_2)$ for Phe (Left column), Tyr (Middle column), and Trp (Right column). Row (**a**): Observed $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp residues in an α -helical backbone conformation. Row (**b**): Calculated $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp dipeptide mimetics with φ and ψ angles within $\pm 10^{\circ}$ of the canonical α -helix values ($\varphi = -57^{\circ}, \psi = -47^{\circ}$). Row (**c**): Calculated $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp in α -helical segments. Row (**d**): Observed $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp residues in a β -sheet backbone conformation. Row (**e**): Calculated $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp in α -helical segments. Row (**d**): Observed $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp residues in a β -sheet backbone conformation. Row (**e**): Calculated $P(\chi_1, \chi_2)$ for Phe, Tyr, and Trp bipeptide mimetics with φ and ψ angles within $\pm 10^{\circ}$ of the canonical β -sheet values ($\varphi = -119^{\circ}, \psi = 113^{\circ}$). The probabilities are normalized such that $\int P(\chi_1, \chi_2) d\chi_1 d\chi_2 = 1$. The probability values, expressed as percentages, are defined by regions between the two vertical dotted lines. The probabilities increase from white to yellow to orange to black. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

for Met dipeptides in α -helix backbone conformations (Table S1 in Supporting Information). Note that the hard-sphere model successfully recapitulates the observed $P(\chi_1)$ and $P(\chi_2)$ distributions for Met, but not $P(\chi_3)$. The observed $P(\chi_3)$ shows strong peaks at $\chi_3 = 60^\circ$ and 300° , whereas the predicted $P(\chi_3)$ is nearly flat over the range $60^\circ < \chi_3 < 300^\circ$.⁴ The difference between the observed and predicted distributions persists when we vary the sulfur atomic size over the range from 1.45 to 2.0Å, artificially place two hydrogen atoms in an sp³ configuration on the sulfur atom, and study Met in the context of an α -helical segment.

We also investigated the amino acid Norleucine (Nle), a structural analog of Met where the thioether group (-S-) is replaced by the methylene group (-CH₂-; Fig. 8). The observed distribution $P(\chi_3)$ for Nle is different from that of Met in that $\chi_3 = 180^\circ$ is the most probable value of χ_3 . In Figure 9, we show that the $P(\chi_1)$, $P(\chi_2)$, and $P(\chi_3)$ distributions from the hard-sphere dipeptide model for Nle agree qualitatively with the observed distributions. (Note that the observed Nle distributions possess large fluctuations due to the small number of Nle in the PDB). In future studies, we will investigate why the hard-sphere plus stereochemical constraint model can recapitulate $P(\chi_3)$ for Nle, but not for Met. To confirm that the observed $P(\chi_3)$ is accurate, we will also investigate the electron density map for each Met residue and consider possible multiple conformations of the Met side chains using Ringer.³⁵

Unexpectedly, we discovered that the hard-sphere plus stereochemical constraint model is also able to recapitulate the behavior of the polar side chains Ser and Thr in



Stick representation of Phe within an α -helical segment. Stick representation of Phe at the central position *i* within a nine-residue α -helical segment and its side-chain dihedral angle χ_1 set to 180° (Left) and 300° (Right). The C^{*i*+4} and C^{*i*-4} atoms are labeled and the atomic clashes that occur between the carbonyl group of residue *i*-4 the side-chain of Phe when $\chi_1 = 300^\circ$ are highlighted by semitransparent spheres. Carbon, nitrogen, oxygen, and hydrogen atoms are colored pink, blue, red, and white, respectively. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

both the α -helix and β -sheet backbone conformations without including hydrogen-bonding interactions in the model. For example, our results show that $\chi_1 = 300^\circ$ for Thr is sterically allowed, whereas $\chi_1 = 60^\circ$ and 180° are disallowed by repulsive steric interactions. The $\chi_1 = 300^\circ$ conformation positions the Thr side-chain to hydrogen bond with the backbone.^{36,37} But even in the absence of



Figure 8

Stick representation of the Met and Nle dipeptide mimetics. The sidechain dihedral angles χ_1 , χ_2 , and χ_3 are defined with positive angles indicated by the arrows. Carbon, nitrogen, oxygen, hydrogen, and sulfur atoms are shaded pink, blue, red, white, and yellow, respectively. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]



Figure 9

Side-chain dihedral angle distributions for Met and Nle dipeptide mimetics. Comparison of the observed (dotted red lines) and calculated (solid blue lines) probability distributions $P(\chi_1)$ (top), $P(\chi_2)$ (middle), and $P(\chi_3)$ (bottom) for the side-chain dihedral angles χ_1 , χ_2 , and χ_3 for Met (left, binned by 5°) and Nle (right, binned by 20°) in dipeptide mimetics with α -helix backbone conformations. The probabilities are normalized such that $\int P(\chi_n) d\chi_n = 1$, with n = 1, 2, or 3. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

H-bonding, the hard sphere model predicts that $\chi_1 = 300^{\circ}$ is the most populated conformation. We have not yet investigated charged side chains (Lys, Arg, Glu, and Asp) or residues with amide side chains (Asn and Gln).

In summary, we found that side-chain dihedral angle distributions of many amino acids can be accurately modeled using hard-sphere interactions plus stereochemical constraints alone. The success of the hard sphere plus stereochemical constraint model in predicting amino acid side-chain conformation is a strong foundation from which to investigate more complex systems and phenomena, such as the packing and thermodynamic stability of protein cores and protein-protein interactions.

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REFERENCES

- 1. Ramakrishnan GN, Ramakrishnan C, Sasisekharan V. Stereochemistry of polypeptide chain configurations. J Mol Biol 1963;7:95–99.
- Ramakrishnan C, Ramachandran GN. Stereochemical criteria for polypeptide and protein chain conformations. II. Allowed conformations for a pair of peptide units. Biophys J 1965;55:909–933.

- Laskowski RA, MacArthur MW, Moss DS, Thornton JM. PRO-CHECK: A program to check the stereochemical quality of protein structures. J Appl Cryst 1993;26:283–291.
- Davis IW, Murray LW, Richardson JS, Richardson DC. MOLPRO-BITY: Structure validation and all-atom contact analysis for nucleic acids and their complexes. Nucleic Acids Res 2004;32:W615–619.
- Dunbrack RL, Jr., Karplus M. Conformational analysis of the backbone-dependent rotamer preferences of protein sidechains. Nat Struct Biol 1994;1:334–340.
- Bower MJ, Cohen FE, Dunbrack RL, Jr. Prediction of protein sidechain rotamers from a backbone-dependent rotamer library: a new homology modeling tool. J Mol Biol 1997;267:1268–1282.
- Dunbrack RL, Jr., Cohen FE. Bayesian statistical anal- ysis of protein side-chain rotamer preferences. Protein Sci 1997;6:1661–1681.
- Shapovalov MV, Dunbrack RL, Jr. A smoothed backbone-dependent rotamer library for proteins derived from adaptive kernel density estimates and regressions. Structure 2011;19:844–858.
- Zhu X, Lopes PEM, Shim J, MacKerell AD. Intrinsic energy landscapes of amino acid side-chains. J Chem Info Modeling 2012;52: 1559–1572.
- Cornell WD, Cieplak P, Bayly CI, Gould IR, Merz KM, Jr., Ferguson DM, Spellmeyer DC, Fox T, Caldwell JW, Kollman PA. A Second Generation Force Field for the Simulation of Proteins. J Am Chem Soc 1995;117:5179–5197.
- Brooks BR, Bruccoleri RE, Olafson BD, States DJ, Swaminathan S, Karplus M. CHARMM: a program for macromolecular energy, minimization, and dynamics calculations. J Comp Chem 1983;4:187– 217.
- Van Der Spoel D, Lindahl E, Hess B, Groenhof G, Mark AE, Berendsen HJ. GROMACS: fast, flexible, and free. J Comput Chem 2005;26:1701–1718.
- Kaminski GA, Friesner RA, Tirado-Rives J, Jorgensen WL. Evaluation and Reparametrization of the OPLS-AA force field for proteins via comparison with accurate quantum chemical calculations on peptides. J Phys Chem B 2001;105:6474–6487.
- MacKerell AD, Jr., Feig M, Brooks CL, III. Extending the treatment of backbone energetics in protein force fields: limitations of gasphase quantum mechanics in reproducing protein conformational distributions in molecular dynamics simulations. J Comput Chem 2004;25:14001415.
- 15. Best RB, Zhu X, Shim J, Lopes PEM, Mittal J, Feig M, MacKerell AD. Optimization of the additive CHARMM all-atom protein force field targeting improved sampling of the backbone φ , ψ and Side-Chain χ_1 and χ_2 dihedral angles. J Chem Theory Comput 2012;8: 3257–3273.
- Lindorff-Larsen K, Piana S, Palmo K, Maragakis P, Klepeis JL, Dror RO, Shaw DE. Improved side-chain torsion potentials for the Amber ff99SB protein force field. Proteins 2010;78:1950– 1958.
- Fleishman SJ, Whitehead TA, Ekiert DC, reyfus C, Corn JE, Strauch EM, Wilson IA, Baker D. Computational design of proteins targeting the conserved stem region of influenza hemagglutinin. Science 2011 332:816–821.
- Wang G, Dunbrack RL, Jr. PISCES: recent improvements to a PDB sequence culling server. Nucleic Acids Res 2005;33:W94–W98.

- Zhou AQ, OHern CS, Regan L. The power of hard-sphere models: explaining side-chain dihedral angle distributions of Thr and Val. Biophys J. 2012;102:2345–2352.
- 20. Zhou AQ, Caballero D, O'Hern CS, Regan L. New insights into the interdependence between amino acid stereochemistry and protein structure. Biophys J 2013;105:2403–2411.
- Zhou AQ, O'Hern CS, Regan L. Revisiting the Ramachandran plot from a new angle. Protein Sci 2011;20:1166–1171.
- 22. Bondi A. van der Waals volumes and radii. J Phys Chem 1964;68: 441–452.
- Element data and radii, Cambridge Crystallographic Data Centre. Available at: http: //www.ccdc.cam.ac.uk/products/csd/radii. Accessed on December 4, 2013.
- 24. Seeliger D, de Groot BL. Atomic contacts in protein structures. A detailed analysis of atomic radii, packing, and overlaps. Proteins 2007;68:595–601.
- Pauling L. The Nature of the Chemical Bond, 2nd ed. Ithaca, NY: Cornell University Press; 1948. pp 187–193.
- 26. Chothia C. Structural invariants in protein folding. Nature 1975; 254:304–308.
- Richards FM. The interpretation of protein structures: total volume, group volume distributions and packing density. J Mol Biol 1974; 82:1–14.
- 28. Li AJ, Nussinov R. A set of van der Waals and Coulombic radii of protein atoms for molecular and solvent-accessible surface calculation, packing evaluation, and docking. Proteins Struct Funct Genet 1998;32:111–127.
- 29. Momany FA, Carruthers LM, Scheraga HA. Intermolecular potentials from crystal data. III Determination of empirical potentials and application to the packing configurations and lattice energies in crystals of hydrocarbons, carboxylic acids, amines and amides. J Phys Chem 1974;78:1595–1630.
- Allinger NL, Yuh YH. Quantum Chemistry Program Exchange, Indiana University, 1980; Program 395.
- Word JM, Lovell SC, Richardson JS, Richardson DC. Asparagine and glutamine: using hydrogen atom contacts in the choice of sidechain amide orientation. J Mol Biol 1999;285:1735–1747.
- 32. Caballero D, Määttä J, Zhou AQ, Sammalkorpi M, O'Hern CS, Regan L. The intrinsic α-helical and β-sheet preferences: a computational case study of Alanine. Prot Sci 2014;23:970.
- Richardson JS. The anatomy and taxonomy of protein structure. Adv Protein Chem 1981;34:167–339.
- Mcgregor MJ, Islam SA, Sternberg MJE. Analysis of the relationship between side-chain conformation and secondary structure in globular-proteins. J Mol Biol 1987;198:295–310.
- 35. Lang PT, Ng HL, Fraser JS, Corn JE, Echols N, Sales M, Holton JM, Alber T. Automated electron-density sampling reveals widespread conformational polymorphism in proteins. Protein Sci 2010;19: 1420–1431.
- McGregor MJ, Islam SA, Sternberg MJE. Analysis of the relationship between side-chain conformation and secondary structure in globular proteins. J Mol Biol 1987;198:295–310.
- 37. Gray TM, Matthews BW. Intrahelical hydrogen bonding of serine, threonine, and cysteine residues within α -helices and its relevance to membrane-bound proteins. J Mol Biol 1984;175:75–81.